

328. *The Constitution of the Purine Nucleosides. Part III. Potentiometric Determination of the Dissociation Constants of Methylated Xanthines.*

By ALEXANDER G. OGSTON.

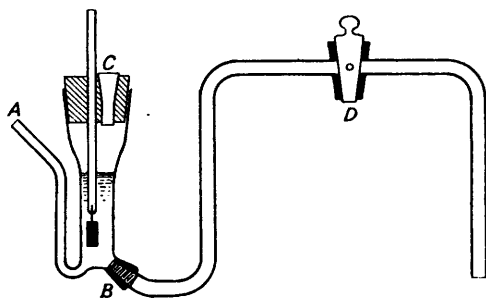
THE measurement of the dissociation constants of methylated xanthines was undertaken at the suggestion of Dr. J. M. Gulland and Mr. E. R. Holiday. It was hoped to combine this evidence with that of organic chemical methods and that of ultra-violet absorption spectra, in order to throw light on their constitution. The previous work on the dissociation constants of this class of substance is not comprehensive; the constants of a few xanthines were determined by Wood (J., 1903, **83**, 568; 1906, **89**, 1831, 1839). The figures now obtained differ in some cases considerably from those of the earlier workers.

The fundamental difficulties in determining dissociation constants by the method of potentiometric titration have been discussed by Ogston and Brown (*Trans. Faraday Soc.*, 1935, **31**, 166). If the liquid junction potential is not affected by the composition of the titrated liquid, the *E.M.F.* of the cell is proportional to the mean activity of the acid solution which it contains; if this solution is sufficiently dilute, the activity coefficients in it will not differ greatly from unity, the *E.M.F.* can be taken as proportional to the concentration of hydrogen ions, and the dissociation constant can be written without great error in its classical form $K = [H][A]/[HA]$, where A and HA are the basic and the acid form of the titrand. For this reason and because of the scarcity and insolubility of some of the materials, the solutions titrated were rarely more concentrated than $M/1000$ and sometimes as dilute as $M/3000$.

EXPERIMENTAL.

The materials used were supplied in a pure form by Dr. J. M. Gulland.

The cell was of the form shown in the figure. Its capacity was 3 c.c.; hydrogen was introduced through the side tube and the bubbles stirred the liquid in the cell very efficiently. The liquid junction was made at the end of the bridge tube, which was closed by a porous plug of sintered glass and ground into a joint at the base of the cell; the bridge was filled with the bridge solution (3.5*N*-potassium chloride) by removing it from the cell and applying suction at the closed end; superfluous liquid was removed from the sintered end before the bridge was replaced in the cell. This type of liquid junction gave results reproducible to 1 mv. when freshly made, and constant to within 3 mv. during several hours; it also allowed the titrand solution to be completely stirred during titration with a minimum disturbance of the junction. Flow of liquid was prevented by the closed unlubricated tap on the bridge tube, which had a resistance so



- A. Hydrogen tube.
- B. Sintered glass plug.
- C. Hydrogen exit and entrance for micro-burette.
- D. Closed ungreased tap.

low that it could be kept closed during measurements.

The reference electrode was 3.5*N*-calomel; the platinum electrodes were prepared and used as described by Ogston and Brown (*loc. cit.*). The p_H potential scale was standardised with $N/10$ -hydrochloric acid, its activity coefficient being taken into account. Measurements were made at $18^\circ \pm 0.5^\circ$ in a constant-temperature room.

Titrand $N/10$ -solutions were added from a capillary microburette: this had an arbitrary scale and was calibrated by weighing water delivered by it. The total capacity was 0.3 c.c. and the accuracy $\pm 10^{-4}$ c.c.

The constants were calculated from the experimental points by the method of Ogston and Brown; the values in water are the mean in each case of two or more concordant runs and are probably accurate to $\pm 0.05 p_H$ unit; those in alcoholic solutions are less accurate owing to the

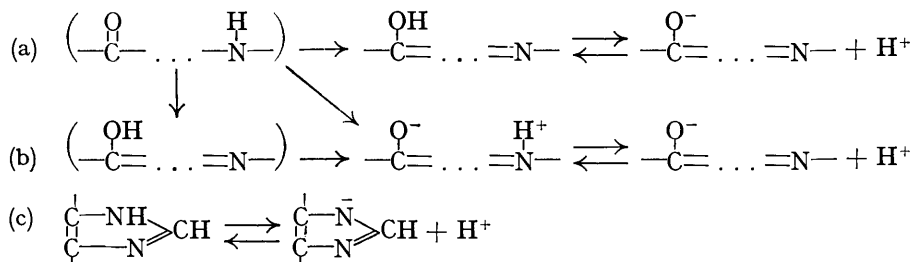
less reproducible behaviour of the electrodes in this solvent, but are probably accurate to ± 0.1 p_K unit.

Substance.	p_K .		Δp_K .	Substance.	p_K .		Δp_K .
	Water.	90% EtOH.			Water.	90% EtOH.	
Xanthine	7.7	9.3	1.6	7-Methylxanthine	8.5	8.8	0.3
1-Methyl-	7.7	9.2	1.5	1:7-Dimethyl- ...	8.5	8.7	0.2
3:7-Dimethyl-	9.9	11.3	1.4	9-Methyl-	6.3	6.8	0.5
3-Methyl-	8.5	8.6	0.1	1:9-Dimethyl-	6.3	6.6	0.3
1:3-Dimethyl-	8.6	8.7	0.1	Xanthosine.....	6.0	6.6	0.6

DISCUSSION.

Most of the xanthines are both acids and bases; the acid dissociation constants only have been measured (Table). The basic functions are in all cases very weak, so the amount of buffering produced by the dilute solutions used was small; in terms of potential, the buffering produced was of the same order of magnitude as the reproducibility of the liquid junction potential, so no accurate estimate of their constants could be made. The acid dissociation constants are high enough to be accurately measurable.

Xanthines can conceivably function as acids in three ways: (a) as hydroxy-acids, following enolisation of one of the carbonyl groups; (b) as charged amino-acids, by "Zwitter" ion formation from (a); (c) as glyoxaline acids. These may be represented by the schemes:



The number of possible ways in which xanthine itself might function as an acid is thus considerable, though the number will be smaller for its methyl substituted derivatives. A consideration of the values obtained for the dissociation constants of deoxyxanthines by Tafel and Dodt (*Ber.*, 1907, **40**, 3757) also limits the number of possibilities which need be considered. Their results can be summarised as follows.

(1) Those compounds showing acid properties all have the glyoxaline ring unsubstituted: their dissociation constants vary from 3×10^{-12} to 8×10^{-12} . This class includes the 1:3-dimethyl compound.

(2) All the compounds examined which were substituted in the 7-position, and only these, showed no acid properties.

It must follow from these results that the deoxy-compounds owe their acidity only to ionisation of the $-\text{NH}-$ in the glyoxaline ring; the 2-carbonyl group seems not to be capable of enolisation. It is therefore probably safe to infer that in the xanthines, acid properties must be due either to ionisation of the glyoxaline $-\text{NH}-$ or to enolisation of the 6-carbonyl group; but the great difference between the values of the dissociation constants of the xanthines here obtained and those of the deoxyxanthines, as well as the failure of 7-substitution to destroy the acid properties of xanthines, makes it seem improbable that the acidity of the xanthines is due to ionisation of the glyoxaline $-\text{NH}-$ group. It follows that the acid properties of the xanthines may be ascribed to simple enolisation or "Zwitter" ion formation involving the 6-carbonyl group. On this basis we may proceed to discuss the significance of the present results.

It is possible to distinguish between acids of types (a) and (b) by observing the change of p_K between water and 90% alcoholic solutions (see Linderström-Lang, *Compt. rend. Lab. Carlsberg*, 1927, **4**, 6; Neuberger, *Proc. Roy. Soc.*, 1934, **B**, **115**, 180; and Ogston and Brown, *loc. cit.*). Uncharged acids of type (a) give a comparatively large positive

